SERION ELISA classic Parainfluenza Virus IgA

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SERION ELISA classic Parainfluenza Virus IgA

Enzyme immunoassay for determination of human antibodies For sale in the U.S. for Research Use Only. Not for diagnostic use.

SERION ELISA *classic* Parainfluenza Virus IgA

Order no. ESR126A

1 INTENDED USE

The SERION ELISA classic Parainfluenza Virus IgA test is a qualitative and quantitative immunoassay for the detection of human antibodies in serum or plasma directed against all relevant human pathogenic Parainfluenza Viruses.

For sale in the U.S. for Research Use Only. Not for diagnostic use.

2 BACKGROUND

Parainfluenza Viruses cause mild to severe upper and lower respiratory tract infections. Parainfluenza Viruses are surpassed only by Respiratory Syncytial Virus as the cause of severe lower respiratory tract infections in children. Infections are transmitted by droplet infection or direct contact via mucous membranes of the eyes, mouth, or nose. Currently, Parainfluenza Viruses can be divided into four type of which types 1-3 are most frequently detected.

3 TEST PRINCIPLE SERION ELISA classic

The ELISA (Enzyme-Linked Immunosorbent Assay) is an immunoassay suited to the detection of antibodies. The reaction is based on the specific interaction of antibodies with their corresponding antigen. The test strips of the SERION ELISA *classic* microtiter plate are coated with specific antigens of the pathogen of interest. If antibody in a sample is present, they bind to the fixed antigen. A secondary antibody, which has been conjugated with the enzyme alkaline phosphatase, detects and binds to the antigen-antibody complex. The colorless substrate p-nitrophenylphosphate is then converted into the colored product p-nitrophenol. The signal intensity of this reaction product is proportional to the concentration of the antibody in the sample and is measured photometrically.

4 KIT COMPONENTS

| Test Components | Pieces/ Volume |
|-------------------------------------------------------------------------------------------------------------------------------------------|-------------------|
| Production of the feet string each with sight entires a set of single wells | |
| Break apart microtiter test strips each with eight antigen coated single wells, | 12 pieces |
| (altogether 96) MTP, 1 frame. The coating material is inactivated. | |
| Standard serum (ready-to-use) STD, | 2 x 2 ml |
| Human serum in protein-containing phosphate buffer; negative for anti-HIV Ab, HBs-Ag (Hepatitis B-Virus surface Antigen) and anti-HCV Ab; | |
| Preservative: <0.1% sodium azide; coloring: Amaranth O | |
| Negative control serum (ready-to-use) NEG, | 2 ml |
| Human serum in protein-containing phosphate buffer; negative for anti-HIV Ab, HBs-Ag (Hepatitis B-Virus surface Antigen) and anti-HCV Ab; | |
| Preservative: <0.1% sodium azide; coloring: Lissamin Green V | |
| Anti-human IgA conjugate (ready-to-use) APC | 13ml |
| Anti-human IgA, IgG or IgM polyclonal antibody, | |
| Conjugated to alkaline phosphatase, stabilized with protein stabilization solution; | |
| Preservative: <0.1% methylisothiazolone, <0.1% bromnitrodioxane | |
| Washing solution concentrate (sufficient for 1000ml WASH, | 33.3ml |
| Sodium chloride solution with Tween 20 and 30mM Tris-HCl, pH 7.4; | |
| Preservative: <0.1% sodium azide | |
| Dilution buffer (ready-to-use) DILB, | 2 x 50ml |
| Protein-containing phosphate buffer with Tween 20; | |
| Preservative: <0.1% sodium azide; coloring: 0.01g/l Bromphenol blue | |
| Stopping solution (ready-to-use) STOP, | 15ml |
| <0.1N sodium hydroxide, 40mM EDTA | |
| Substrate (ready-to-use) pNPP, | 13ml |
| Para-nitrophenylphosphate in solvent-free buffer; | |
| Preservative: <0.1% sodium azide | |
| Quality control certificate with standard curve and evaluation table INFO, | 2 pages |
| (quantification of antibodies in IU/ml or U/ml) | |

5 MATERIAL REQUIRED BUT NOT SUPPLIED

- Common laboratory equipment
- Photometer for microtiter plates with filter, wavelength 405nm, recommended reference wavelength 620nm-690nm (e.g., 650nm)
- Microtiter plate washer
- Incubator 37°C
- Moist chamber
- Distilled water
- Optional: SERION ELISA control

6 STORAGE AND STABILITY

| Reagent | Storage | Stability |
|---------------------------------|--------------------------------------------------------------------------------------------------|--------------------------------------|
| Microtiter strips (coated with | Unopened | See expiry date |
| antigen) | After opening at 2-8°C in closed aluminum bag with desiccant | Minimum shelf-life: four weeks |
| Control sera / Standard sera | Unopened / after opening at 2-8°C | See expiry date |
| Conjugate | Unopened / after opening at 2-8°C | See expiry date |
| Dilution buffer | Unopened / after opening at 2-8°C | See expiry date |
| Washing solution | Unopened / after opening at 2-8°C Working dilution at 2-8°C Working dilution at room temperature | See expiry date 2 weeks 1 week |
| Substrate | Unopened / after opening at 2-8°C | See expiry date |
| Stopping solution | Unopened / after opening at 2-8°C | See expiry date |

7 TEST PROCEDURE SERION ELISA classic

7.1 Evidence of Deterioration

Optimum results can only be achieved if the instructions are strictly followed. Only use SERION ELISA *classic* reagents when using SERION ELISA *classic* immunoassays. The components must not be exchanged for reagents of other manufacturers. Standard and control sera of SERION ELISA *classic* immunoassays are defined exclusively for the test kit to be used and must not be used in other lots. Washing solution, substrate and stop solution can be used for all SERION ELISA *classic* immunoassays irrespective of the lot and the test.

Each SERION ELISA *classic* test contains a ready-to-use sample dilution buffer. In some cases the use of special dilution buffers is necessary to guarantee consistent quality and reliable results. The dilution buffers can be used irrespective of the lots.

There are three different conjugate concentrations for each immunoglobulin class (IgA, IgG, IgM) indicated on the label as + (low), ++ (medium), and +++ (high). Conjugates with the same concentration and of the same immunoglobulin class are interchangeable and can be used for other SERION ELISA *classic* immunoassays irrespective of the lot and the test. Dilution or alteration of the reagents may result in a loss of sensitivity. Use aseptic techniques when removing aliquots from the reagent tubes to avoid contamination.

Reproducibility of test results is dependent on thorough mixing of the reagents. Agitate the vials containing control sera before use and also all samples after dilution (e.g., by using a vortex mixer).

Be sure to pipette carefully and comply with the given incubation times and temperatures. Significant time differences between pipetting the first and last well of the microtiter plate when dispensing samples and control sera, conjugate or substrate can result in different pre-incubation times, which may influence the precision and reproducibility of the results. Avoid exposure of reagents to strong light during storage and incubation.

Adequate washing avoids test unspecificities. Therefore, the washing procedure should be carried out carefully. All of the flat bottom wells should be filled with equal volumes of washing buffer. At the end of the procedure ensure that the wells are free of all washing buffer in order to avoid uncontrolled dilution effects. Avoid foaming!

Reagents must be tightly closed after use to avoid evaporation and contamination. Take are not to mix up the caps of the bottles and/or vials.

The SERION ELISA *classic* immunoassay is only valid if the lot-specific validation criteria on the quality control certificate are fulfilled.

7.2 Sample Preparation and Storage

Lipaemic, hemolytic or icteric samples (serum or plasma) should only be tested with caution. Obviously contaminated samples should not be tested. Serum or plasma (EDTA, citrate, heparin) collected according to standard laboratory methods are suitable samples. Samples must not be thermally inactivated.

7.2.1 Dilution of Samples

Before running the test, samples (V_1) must be diluted in dilution buffer (V_2) as follows:

SERION ELISA classic Parainfluenza Virus IgA

| $V_1 + V_2 = 1:400$ | add | 10ul | sample |
|---------------------|---------|---------------|---------------------------------------------------------|
| | each to | 1000ul | dilution buffer (= 1:100) |
| | each to | 50ul 200ul | from the first dilution step dilution buffer (= 1:4) |

After dilution and before pipetting into the microtiter plate, the samples must be mixed thoroughly to prepare a homogenous solution.

7.2.2 Sample Storage

Samples should not be stored for more than 7 days at 2-8°C. Extended storage is possible at ≤-20°C. Avoid repeated freezing and thawing of samples. Diluted samples can be stored at 2-8°C for one week.

7.3 Preparation of Kit Reagents

Bring all reagents to room temperature before testing.

7.3.1 Microtiter Test Strips

The microtiter test strips labeled with abbreviations for pathogen and immunoglobulin class are packed with a desiccant in an aluminum bag. To open the aluminum bag of the microtiter plate, please cut off the top of the marked side only in order to guarantee proper resealing. Take unrequired wells out of the frame and put them back into the aluminum bag. Close bag carefully to ensure airtight conditions. Do not use strips if the aluminum bag is damaged or if the bag with remaining strips and desiccant was not properly resealed.

7.3.2 Control Sera / Standard Sera (ready-to-use)

Control and standard sera are ready-to-use and must not be diluted any further. For each test run (independent of the number of microtiter test strips to be used) control and standard sera must be included. Standard and cut-off sera should be set up in duplicate. Do not treat control sera with Rf-absorbent.

7.3.3 Anti-human IgA AP-Conjugate (ready-to-use)

The required conjugate concentration (i.e., +, ++, +++) is indicated on the quality control certificate. Please refer also to the specification on the label.

7.3.4 Washing Solution (Concentrate)

Dilute washing buffer concentrate (V_1) 1:30 with distilled H2O to a final volume of V_2 . Example:

| Buffer concentrate (V ₁) | Final volume (V ₂) |
|--------------------------------------|--------------------------------|
| 33.3ml | 1000ml |
| 1.0ml | 30ml |

7.3.5 Dilution Buffer for Samples (ready-to-use)

7.3.6 Substrate (ready-to-use)

Substrate in unopened bottle may have a slight yellow color which does not reduce the quality of the product!

7.3.7 Stopping Solution (ready-to-use)

7.4 **Overview - Test Procedure**

SERION ELISA classic Parainfluenza Virus IgA quantitative

sample dilution¹ IgA 1:500

Pipette diluted samples and ready-to-use control/ standard sera into the microtiter wells (100ul/well)

INCUBATION 60min/37°C moist chamber WASH (4 x 300ul/well DIL WASH)² Pipette conjugate solution APC (100ul/well)

INCUBATION 30min/37°C moist chamber

WASH (4 x 300ul/well DIL WASH)²

Pipette substrate solution pNPP (100ul/well)

INCUBATION 30min/37°C moist chamber

Pipette stopping solution STOP (100ul/well)

READ EXTINCTION at 405nm

¹Special dilution buffers for the following SERION ELISA classic tests: Borrelia burgdorferi IgG, IgM, EBV EA IgG and Hantavirus Puumala IgG, IgM

²For manual use: tap plate at the end of the wash procedure on paper towel.

7.5 Manual Test Procedure

- 1. Place the required number of **wells in the frame** and prepare a protocol sheet.
- 2. Add each **100ul of diluted sample or ready-to-use controls** into the appropriate wells of microtiter test strips. Spare one well for substrate blank, e.g.:

| Well | Quantitative ELISA |
|------|--------------------|
| A1 | Substrate blank |
| B1 | Negative control |
| C1 | Standard serum |
| D1 | Standard serum |
| E1 | Sample 1 |
| F1 | Sample 2 |

- 3. **Sample incubation** for 60 minutes (+/- 5 min) at 37°C (+/- 1°C) in moist chamber.
- 4. After incubation **wash** all wells with washing solution (by automated washer or manually):
 - aspirate or shake out the incubation solution
 - fill each well with 300ul washing solution
 - aspirate or shake out the washing solution
 - repeat the washing procedure 3 times (altogether 4 times!)
 - dry by tapping the microtiter plate on a paper towel

5. Addition of conjugate

Add 100ul of the ready-to-use IgA conjugate to the appropriate wells (except substrate blank).

- 6. **Conjugate incubation** for 30 minues (+/- 1 min) at 37°C (+/- 1°C) in moist chamber.
- 7. After incubation **wash** all wells with washing solution (see above).
- 8. Addition of substrate

Add 100ul of ready-to-use substrate solution to each well (including well for substrate blank!)

- 9. **Substrate incubation** for 30 minutes (+/- 1 min) at 37°C (+/- 1°C) in moist chamber.
- 10. Stopping the reaction

Add 100ul of stopping solution to each well, shake microtiter plate gently to mix.

11. Read extinction

Read optical density (OD) within 60 minutes at 405nm against substrate blank, reference wavelength between 620nm and 690nm (e.g. 650nm).

7.6 Automated Test Procedure

SERION ELISA are suited for processing on automats and evaluated for use with Immunomat[™] and Gemini as well as with DYNEX DSX® and DS2®. The automated processing is performed analogous to manual use. Please note that under special working conditions (e.g. ambient temperature) internal laboratory adaptations of the substrate incubation times may be necessary.

7.7 Positive Control / Accuracy Control

For the periodic verification of the test method, in order to fulfill the requirements of laboratory internal quality management systems, we recommend using SERION ELISA *controls* to determine precision and accuracy of SERION ELISA *classic* test runs. The use of SERION ELISA *controls* is described in specific instruction manuals.

8 TEST EVALUATION

8.1 SERION ELISA classic Parainfluenza Virus IgA

The mathematical curve fitting for antibody quantification with SERION ELISA classic immunoassays is based on the 4-parameter logistic (4 PL) function.

Activity
$$(U/ml) = e^{C - \frac{1}{B} \ln(\frac{D-A}{OD(Patient)*F-A} - 1)}$$

The 4 parameters A, B, C, and D are representative for the exact shape of the standard curve:

Parameter A: Lower asymptote (OD)
Parameter B: Slope of the curve
Parameter C: Inflection point

Parameter D: Upper asymptote (OD)

Institut Virion\Serion GmbH establishes a lot-specific 4 PL standard curve for each SERION ELISA *classic* immunoassay in multiple test runs under optimal test conditions. The four parameters are indicated on the quality control certificate of each individual SERION ELISA *classic* test.

For the adaptation of the test level to the given 4 PL standard curve, the correction factor F is calculated by dividing the standard reference OD value indicated on the quality control certificate with the measured, and consequently test run-specific, standard OD value.

$$F = \frac{STD \ reference \ OD \ value}{measured \ STD \ OD \ value}$$

By multiplying the OD values obtained from samples with the correction factor F, the level of each individual test run is adjusted to the given 4 PL standard curve. Thereby, interassay deviations are compensated for and antibody activities can be directly evaluated from the 4 PL standard curve.

After subtraction of the substrate blank from all measured OD values and calculation of the mean OD value of the standard serum (STD), tested in duplicate, a range of possibilities are available for the evaluation of antibody activities from the optical measurement signals (OD) of samples. They are described in separate manuals.

8.2 Borderline Ranges

The borderline ranges of the SERION ELISA *classic* Parainfluenza Virus IgA test are specified on the quality control certificates and indicate the range of borderline test results. Values below this range indicate a negative result; values above the borderline range indicate a positive result.

8.3 Limits of Quantification

The limits of quantification are specified on the quality control certificate of the SERION ELISA *classic* Parainfluenza Virus IgA. The linearity of dilution within this range has been demonstrated in comprehensive evaluation studies. If a sample shows a test result above the

upper limit of quantification, the sample may be tested at a higher dilution. The resulting antibody activity must then be multiplied by the additional dilution factor.

8.4 Automated Evaluation / Software

For the automated evaluation of optical measurement signals, the software SERION easy *ANALYZE*, the software SERION *evaluate* as well as the Microsoft® Excel®-based software tool SERION *activity* are available on request.

8.5 Criteria of Validity

- The substrate blank must be <0.25 OD.
- The negative control must be negative.
- By use of quantitative SERION ELISA *classic* tests, the mean OD value (after subtraction of the substrate blank!) of the standard serum must be within the validity range which is given on the lot-specific quality control certificate.
- By use of qualitative SERION ELISA *classic* tests, the OD value of the positive control and the mean OD value of the cut-off serum must be within the validity ranges which are given on the lot-specific quality control certificate of the kit (after subtraction of the substrate blank!).
- The variation of OD values of the standard serum or cut-off serum must not be higher than 20%.

If these criteria are not met, the test is not valid and must be repeated.

9 SAFETY MEASURES

9.1 Statements of Warning

The SERION ELISA *classic* is designed for use by qualified personnel who are familiar with good laboratory practice. All kit reagents and human samples should be handled carefully using established good laboratory practice.

- This kit contains human blood components. Although all control- and cut-off sera have been tested and found negative for anti-HIV Ab, HBs-Ag (*Hepatitis B Virus surface Antigen*) and anti-HCV Ab, they should be considered potentially infectious.
- Do not pipette by mouth.
- Do not smoke, eat, or drink in areas in which samples or kit reagents are handled.
- Wear disposable gloves, laboratory coat, and safety glasses while handling kit reagents or samples. Wash hands thoroughly afterwards.
- Samples and other potentially infectious material should be decontaminated after use.
- Reagents should be stored safely and be inaccessible to unauthorized access, e.g. children.

9.2 Disposal

Please observe the relevant statutory requirements!