





Product No. 7305
recomWell SARS-CoV-2 IgA

## 1 Purpose

For sale for Research Use Only. Not for diagnostic use.

### 2 Intended Use

MIKROGEN's *recom*Well SARS-Cov-2 IgA detects IgA antibodies against SARS-CoV-2 (severe acute respiratory syndrome coronavirus-2). **This test must not be used for clinical diagnosis!** 

### 3 Test Principle

Highly purified recombinant nucleocapsid protein of SARS-CoV-2 is fixed to the wells of a microplate.

- Diluted serum or plasma samples are incubated in the wells; antibodies in the samples bind specifically to the antigen on the wells.
- 2. Unbound antibodies are washed away.
- 3. In a second step, anti-Human Immunoglobulin antibody, coupled to horseradish peroxidase (HRP) is incubated in the wells.
- 4. Unbound conjugate is washed away.
- 5. Specifically-bound antibodies are detected by a peroxidase-catalyzed color reaction. Where an antigen-antibody reaction has taken place, the color development of the chromogenic substrate is proportional to the quantity of bound SARS-CoV-2 IgA antibodies. The concentration of color development can be measured using a photometer so as to establish the concentration of anti-SARS-CoV-2 IgA antibodies in the samples.

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### 4 Reagents

## 4.1 Package contents

The reagents in one package are sufficient for 96 tests. Each test kit contains:

<b>12 x 8</b> wells microplate coated with recombinant SARS-CoV-2 antigens in a vacuum pressure-
I SARS-CoV-2 antigens in a vacuum pressure-
<del></del>
sealed bag
100ml Wash buffer (ten times concentration)
WASHBUF 10X Contains phosphate buffer, NaCl and detergent
Preservative: MIT (0.01%) and Oxypyrion (0.1%)
125ml Dilution buffer (ready-to-use)
<b>DILUBUF</b> Contains protein, detergent and blue dye
Preservative: MIT (0.01%) and Oxypyrion (0.1%)
SUBS-TMB 12ml Chromogenic substrate
tetramethylbenzidine (TMB, ready-to-use)
SOLN-STOP 12ml stop solution 24.9% phosphoric acid
(H <sub>3</sub> PO <sub>4</sub> )(ready-to-use)
CONTROL + IgA 450ul positive control (brown cap), contains MIT
(0.1%) and Oxypyrion (0.1%)
<b>CONTROL ± IgA</b> 450ul cutoff control (orange cap), contains MIT
(0.1%) and Oxypyrion (0.1%)
CONTROL – IgA 450ul negative control (white cap), contains MIT
(0.1%) and Oxypyrion (0.1%)
CONJ-IgA 500ul anti-human IgA conjugate (101-times
concentrated, blue cap) contains NaN3
(<0.1%), MIT (<0.01% and chlorazetamide
(<0.1%)
(<0.170)
INSTRU 1 Instructions for use

# 4.2 Materials required but not supplied

- Deionized water (high quality)
- Test tubes
- Vortex mixer or other rotators
- o 8-channel pipette or washer with pump
- o Clean measuring cylinders, 50ml and 1000ml
- o Micropipettes with single-use tips, 10ul and 1000ul
- o 10ml pipette or dispenser
- o Incubation chamber 37°C
- o Microplate photometer
- o Timer
- Disposable protective gloves
- Waste container for biohazardous materials

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### 5 Shelf life and handling

- Store reagents at +2°C to +8°C before and after use, do not freeze.
- Before starting the test, allow all reagents to sit for at least 30 minutes at room temperature (+18°C to +25°C).
- The dilution buffer, wash buffer, substrate, and stop solution for the *recom*Well tests can be used for different parameters and batches. At the same time, the shelf life of these components is to be noted.
- The control sera and conjugates are batch-specific and must not be used for different parameters or batches.
- Before use, mix the concentrated conjugates, control reagents and samples thoroughly. Avoid foam formation
- All MIKROGEN microplates are provided with break-apart strips.
- The sealing films are intended for single use only.
- All packages are marked with an expiration date after which quality can no longer be guaranteed.
- Protect kit components from direct sunlight throughout the entire test procedure. The substrate solution (TMB) is especially sensitive to light.
- The test must only be carried out by trained, qualified and authorized personnel.
- Substantial changes made by the user to the product or the directions for use may compromise the intended purpose of the test specified by MIKROGEN.
- Cross-contamination of samples or conjugates in the kit can lead to inaccurate test results. Add
  the samples and conjugate solution carefully. Make sure that incubation solutions do not flow
  over into other wells.
- Automation is possible; further information can be obtained from MIKROGEN.

## 6 Warnings and Precautions

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- All blood products must be treated as potentially infectious.
- The microplate wells have been coated with inactivated whole cell lysates, bacterial, or viral antigens.
- After the addition of samples, the microplate wells must be considered potentially infectious and treated accordingly.
- For the production of control reagents, blood from donors is used which does not contain
  antibodies to HIV-1, HIV-2, HCV, and HBs antigen. These reagents must be treated with the
  same care as experimental samples.
- Suitable single-use gloves must be worn throughout the entire test procedure.
- The conjugates and the wash and dilution buffers contain the antimicrobial agents and preservatives sodium azide (NaN<sub>3</sub>), MIT (methylisothiazolone), oxypyrion, and chloroazetamide. Avoid contact with skin or mucous membranes. Sodium azide can form an explosive azide upon contact with heavy metals such as copper and lead.
- Phosphoric acid is an irritant. Use caution and avoid contact with skin and mucous membranes.
- All fluids to be disposed must be collected. All collection containers must contain suitable
  disinfectants for the inactivation of human pathogens. All reagents and materials contaminated
  with potentially infectious samples must be treated with disinfectants or disposed of according to

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the laboratory guidelines. The concentrations and incubation periods stated by the manufacturer must be observed.

- Only use microplate wells once.
- Do not replace or mix the reagents with reagents from other manufacturers.
- Read through the entire instructions for use before carrying out the test and carefully follow them.
   Deviation from the test protocol described in the instructions for use can lead to erroneous results.

### 7 Sampling and Preparation

### 7.1 Samples

The samples can be serum or plasma (EDTA, citrate, heparin, CPD) and must be separated from as soon as possible after blood collection so as to avoid hemolysis. Avoid microbial contamination of the samples. Insoluble substances must be removed from the samples prior to incubation. The use of heat-inactivated, icteric, hemolytic, lipemic, or turbid samples is not recommended.

CAUTION! If the tests are not carried out immediately, the samples can be stored for up to 2 weeks at +2°C to +8°C. Prolonged storage of the samples is possible at -20°C or less. Repeated freezing and thawing of samples is not recommended due to the risk of producing inaccurate results. Avoid more than 3 cycles of freezing and thawing.

### 7.2 Preparation of solutions

Contains sufficient reagents for 96 tests. The quantities specified below apply to the processing of a single microplate strip with 8 wells. If using several microplate strips at the same time, the specified quantities must be multiplied by the number of microplate strips used. Device-specific dead volume must be taken into account. Dilution buffer, substrate, and stop solution are ready-to-use.

### 7.2.1 Preparation of ready-to-use wash buffer

The wash buffer concentrate is diluted 1 + 9 with  $H_2O$  (deionized water). For each 8-well microplate strip, 5ml concentrate are mixed with 45ml  $H_2O$  (deionized water). The ready-to-use wash buffer can be stored for 4 weeks at +2°C to +8°C or a week at room temperature.

## 7.2.2 Preparation of conjugate solution

For each microplate strip with 8 wells, 1ml of dilution buffer and 10ul of anti-human IgA-peroxidase conjugate (blue cap) are transferred to a clean container and mixed well (dilution = **1+100**). The conjugate solution must be prepared <u>just before use</u>. The ready-to use conjugate solution must not be stored.

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# 8 Test procedure

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No. 1	Expose all reagents for at least 30 minutes to	Note
1	+18°C to +25°C (room temperature) before beginning the test.	Before opening, bring the vacuum-sealed pouch (containing plate) to room temperature to avoid condensation forming in the wells. Remove the required number of strips, re-seal the plate in the pouch and store it in a refrigerator.
		Before use, the control sera and experimental samples as well as the concentrated conjugate must be mixed thoroughly and then briefly centrifuged if possible to collect any liquid on the base of the tubes.
2	Preparing samples and controls Pipette 10ul each of sample and/or control into 1ml dilution buffer and mix well (dilution = 1:100)	Samples and controls must always be diluted immediately prior to carrying out the test.  All controls must be run with each assay.
3	Incubation of samples Pipette 100ul of diluted sample and/or diluted control into each well and incubate for 1 hour at +37°C.	Assign at least one well to the negative control, positive control, and experimental samples. The cutoff (±) control must be assigned to two wells. Preferably, a cutoff control should be included at the beginning of the series and at the end of the series.  For manual processing, carefully cover tightly the microplate with fresh sealing film. Use the incubation chamber at +37°C.
4	Washing	It is recommended to carry out this step with an appropriate ELISA wash device. It is mandatory to ensure that the wash buffer is completely removed between the washing steps.
a) b)	Carefully remove the sealing film. Remove all fluid from the wells.	Aspirate or shake out and tap.
c)	Fill each of the wells with 300ul of ready-to-use wash buffer (see 7.2.1) $\rightarrow$ (8.4b)	Carry out the washing steps 8.4b and 8.4c <b>four times</b> in
5	Incubation with conjugate Add 100ul of diluted conjugate solution ( see 7.2.2) and incubate for 30 minutes at +37°C.	total.  For manual processing, the microplate is carefully covered tightly with unused cover film.
6	Washing (see 8.4b and 8.4c)	Carry out the washing steps four times in total.
7	Substrate reaction Pipette 100ul of ready-to-use substrate solution into each well and incubate for 30 minutes at room temperature. The time is calculated from the time substrate solution is pipetted into the first well.	It is <u>not</u> required to tightly cover the plate. Protect from direct exposure to sunlight.
8	Stopping the reaction Pipette <b>100ul</b> of ready-to-use stop solution into each well.	The substrate solution is not to be removed before adding the stop solution! The same pipetting scheme is to be followed as for the substrate solution.
9	Measurement of the absorbance values The absorbance values of the single wells are measured in a microplate photometer at 450nm and the reference wavelength 620nm (620 to 650nm permitted).	Zero adjustment is done against an air blank. The measurement must be made within 60 minutes of stopping the reaction.

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### 9 Results

### 9.1 Evaluation

Cutoff (threshold) = the arithmetic mean is calculated from the absorbance values of the two cutoff controls (at the start and at the end of the series).

### 9.1.1 Qualitative evaluation

Grey area	Lower limit = cutoff
	Upper limit = cutoff + 20% (cutoff x 1.2)
Negative	Samples with absorbance values <b>below</b> the grey area
Borderline	Samples with absorbance values within the grey area
Positive	Samples with absorbance values above the grey area

### 9.1.2 Quantitative evaluation

The corresponding antibody activity in **units per ml** is assigned to the absorbance values using a formula. The measurement units U/ml are arbitrary units which do not allow conclusions concerning (international) reference values.

U/ml sample	ml sample (Absorbance of sample / absorbance of cutoff) x 20		
Grey area	Lower limit = 20 U/ml		
-	Upper limit = 24 U/ml		
Negative	U/ml sample <20		
Borderline	20 ≤ U/ml sample ≤ 24		
Positive	Positive U/ml sample ≥ 24		

### 9.2 Validation – Quality Control

The test can be evaluated under the following conditions:

- The single absorbance values for the duplicate measurement of the cutoff control do not deviate from their mean by more than 20%.
- Absorbance of the negative control ≤ 0.150.
- Absorbance of the cutoff control absorbance of the negative control ≥ 0.050.
- Absorbance of the positive control absorbance of the cutoff control ≥ 0.300.

The controls are used to validate the test results as per the "Validation Quality Control" section. The reproducibility of results can be improved by determining the specific antibodies relative to the cutoff control in U/ml as the fluctuations caused by carrying out the test are integrated. In validating the test, the positive and negative checks do not need to be evaluated. If necessary, however, they can be carried out for internal quality control purposes. In this case, the results should lie within the target value range given in the analysis certificate or on the label.

### 10 Literature

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- 2. F. Amanat, T. H.O. Nguyen, V. Chromikova, S. Strohmeier, D. Stadlbauer, A. Javier, K. Jiang, G. A. Arunkumar, J. Polanco, M. Bermudez-Gonzales, D. Cap-livski, A. Cheng, K. Kedzierska, O. Vapalahti, J. M. Hepojoki, V. Simon, F. Krammer. A serological assay to detect SARS-CoV-2 seroconversion in humans. Preprint

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# 11 Explanation of symbols

$\sum$	Content is sufficient for <n> applications Number of applications</n>
WASHBUF 10 X	Wash Buffer (ten times concentration)
DILUBUF	Dilution Buffer
SUBS TMB	Chromogenic substrate tetramethylbenzidine
SOLN STOP	Stop solution
TAPE	Covering films
MTP	Microtitre plate
CONTROL + IgA	Positive control IgA
CONTROL ± IgA	Cutoff control IgA
CONTROL - IgA	Negative control IgA
CONJ IgA	Anti-human IgA conjugate
TVALUE	Target and / or target range in U/ml
EVALFORM	Evaluation form
INSTRU	Instructions for use
	See instructions for use
CONT	Contents, includes
RUO	Research use only
LOT	Batch number
X	Do not freeze
REF	Order number
≥	Use by Expiry date
xc \ Yc	Store at x°C to y°C
<b>—</b>	Manufacturer

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## 12 Manufacturer and version information



## 14 Manufacturer and version data

recomWell SARS-CoV-2 IgG			Article no. 7304
recomWel	I SARS-CoV-2 IgA	Article no. 7305	
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