SERION ELISA classic Respiratory Syncytial Virus (RSV) IgA/IgG/IgM

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1 INTENDED USE

The SERION ELISA *classic* Respiratory Syncytial Virus (RSV) IgA/IgG/IgM tests are immunoassays for detection of human antibodies in serum or plasma against Respiratory Syncytial Virus.

For sale in the U.S. for Research Use Only. Not for use in diagnostic procedures.

2 BACKGROUND

Respiratory Syncytial Virus (RSV) causes upper and lower respiratory tract infections predominantly in infants and children. Such infections are responsible for bronchitis as well as pneumonia and may be life threatening. In addition, an RSV infection may often result in middle ear inflammation. In older children and adults, an infection is usually associated with mild symptoms although the elderly may develop more severe symptoms.

RSV is a pleomorphic RNA virus with a diameter of around 150-300nm. Originally isolated from chimpanzees, the virus acquired its name from the ability to form syncytia when cell cultures are infected with the virus.

Infections are transmitted from host to host through contact with mucous membranes of the eyes, mouth, or nose. Epidemiological studies have shown that all children by the age of 2 years have already experienced an RSV infection. Immunity developed as a result of infection is poor due to the virus spreading by syncytia formation. A protective immunity is primarily due to antibody responses to the viral surface glycoproteins G and F, whereby the F protein stimulates a humoral as well as a cellular immune response.

3 TEST PRINCIPLE SERION ELISA *classic*

THE ELISA (Enzyme Linked Immunosorbent Assay) is an immunoassay that is particularly suited to the detection of antibodies. The reaction is based on the specific interaction of antibodies with their corresponding antigen. The test strips of the SERION ELISA *classic* microtiter plate are coated with specific antigens of the pathogen of interest. If antibodies are present in a sample, they bind to the fixed antigen. A secondary antibody that has been conjugated with the enzyme alkaline phosphatase detects and binds to the antigen-antibody complex. The colorless substrate p-nitrophenylphosphate is then converted into the colored product p-nitrophenol. The signal intensity of this reaction product is proportional to the concentration of antibody in the sample and is measured photometrically.
### 4 KIT COMPONENTS

#### Test components

<table>
<thead>
<tr>
<th>Test components</th>
<th>Pieces / Volume</th>
</tr>
</thead>
<tbody>
<tr>
<td>Break apart microtiter test strips each with 8 antigen coated single wells,</td>
<td></td>
</tr>
<tr>
<td>(altogether 96) MTP, 1 frame. The coating material is inactivated.</td>
<td>12 pieces 12</td>
</tr>
<tr>
<td>Standard serum (ready-to-use) STD</td>
<td>2 x 2ml -</td>
</tr>
<tr>
<td>Human serum in protein-containing phosphate buffer; negative for anti-HIV-Ab,</td>
<td></td>
</tr>
<tr>
<td>HBs-Ag (Hepatitis B-Virus surface antigen) and anti-HCV-Ab;</td>
<td></td>
</tr>
<tr>
<td>preservative: &lt;0.1% sodium azide; coloring: Amaranth O</td>
<td></td>
</tr>
<tr>
<td>Cut-off serum (ready-to-use) C/O</td>
<td>2 x 2ml</td>
</tr>
<tr>
<td>Human serum in protein-containing phosphate buffer; negative for anti-HIV Ab,</td>
<td></td>
</tr>
<tr>
<td>HBs-Ag (Hepatitis B-Virus surface antigen) and anti-HCV Ab;</td>
<td></td>
</tr>
<tr>
<td>preservative: &lt;0.1% sodium azide; coloring: Chinaldin yellow</td>
<td></td>
</tr>
<tr>
<td>Positive control serum (ready-to-use) POS</td>
<td>- 2ml</td>
</tr>
<tr>
<td>Human serum in protein-containing phosphate buffer; negative for anti-HIV Ab,</td>
<td></td>
</tr>
<tr>
<td>HBs-Ag (Hepatitis B-Virus surface antigen) and anti-HCV Ab;</td>
<td></td>
</tr>
<tr>
<td>preservative: &lt;0.1% sodium azide; coloring: Amaranth O</td>
<td></td>
</tr>
<tr>
<td>Negative control serum (ready-to-use) NEG</td>
<td>2ml 2ml</td>
</tr>
<tr>
<td>Human serum in protein-containing phosphate buffer; negative for anti-HIV-Ab,</td>
<td></td>
</tr>
<tr>
<td>HBs-Ag (Hepatitis B-Virus surface antigen) and anti-HCV-Ab;</td>
<td></td>
</tr>
<tr>
<td>preservative: &lt;0.1% sodium azide; coloring: Lissamine green V</td>
<td></td>
</tr>
<tr>
<td>Anti-human-IgA, IgG, or IgM conjugate (ready-to-use) APC</td>
<td>13ml 13ml</td>
</tr>
<tr>
<td>Anti-human-IgA, IgG, or IgM polyclonal antibody, conjugated to alkaline</td>
<td></td>
</tr>
<tr>
<td>phosphatase, stabilized with protein stabilization solution;</td>
<td></td>
</tr>
<tr>
<td>preservative: &lt;0.1% methylisothiazolone, &lt;0.1 bromnitrodiolxane</td>
<td></td>
</tr>
<tr>
<td>Washing solution concentrate (sufficient for 1000ml) WASH</td>
<td>33.3ml 33.3ml</td>
</tr>
<tr>
<td>Sodium chloride solution with Tween 20 and 30mM Tris-HCl, pH 7.4;</td>
<td></td>
</tr>
<tr>
<td>preservative: &lt;0.1% sodium azide</td>
<td></td>
</tr>
<tr>
<td>Dilution buffer (ready-to-use) DILB</td>
<td>2 x 50ml 2 x 50ml</td>
</tr>
<tr>
<td>Protein-containing phosphate buffer with Tween 20;</td>
<td></td>
</tr>
<tr>
<td>preservative: &lt;0.1% sodium azide; coloring: 0.01g/l Bromphenol blue</td>
<td></td>
</tr>
<tr>
<td>Stopping solution (ready-to-use) STOP,</td>
<td>15ml 15ml</td>
</tr>
<tr>
<td>&lt;0.1N sodium hydroxide, 40mM EDTA</td>
<td></td>
</tr>
<tr>
<td>Substrate (ready-to-use) pNPP</td>
<td>13ml 13ml</td>
</tr>
<tr>
<td>Para-nitrophenylphosphate in solvent free buffer;</td>
<td></td>
</tr>
<tr>
<td>preservative: &lt;0.1 % sodium azide</td>
<td></td>
</tr>
<tr>
<td>Quality control certificate with standard curve and evaluation table INFO</td>
<td>2 pages -</td>
</tr>
<tr>
<td>(quantification of antibodies in IU/ml or U/ml)</td>
<td></td>
</tr>
<tr>
<td>Quality control certificate INFO</td>
<td>- 1 page</td>
</tr>
</tbody>
</table>

### 5 MATERIAL REQUIRED BUT NOT SUPPLIED

- Common laboratory equipment
- For the IgM detection: SERION Rf-Absorbent (Order no. Z200/20ml)
- Photometer for microtiter plates with filter, wave length 405 nm, recommended reference wave length 620 nm - 690 nm (e.g. 650 nm)
- Microtiter plate washer
- Incubator 37°C
- Moist chamber
- Distilled water
- Optional: SERION ELISA control
6 STORAGE AND STABILITY

<table>
<thead>
<tr>
<th>Reagent</th>
<th>Storage</th>
<th>Stability</th>
</tr>
</thead>
<tbody>
<tr>
<td>Microtiter strips (coated with antigen)</td>
<td>Unopened</td>
<td>See expiry date</td>
</tr>
<tr>
<td></td>
<td>After opening at 2-8°C in closed aluminum-bag with desiccant</td>
<td>Minimum shelf-life: four weeks</td>
</tr>
<tr>
<td>Control sera / Standard sera</td>
<td>Unopened / after opening at 2-8°C</td>
<td>See expiry date</td>
</tr>
<tr>
<td>Conjugate</td>
<td>Unopened / after opening at 2-8°C</td>
<td>See expiry date</td>
</tr>
<tr>
<td>Dilution buffer</td>
<td>Unopened / after opening at 2-8°C</td>
<td>See expiry date</td>
</tr>
<tr>
<td>Washing solution</td>
<td>Unopened / after opening at 2-8°C Working dilution at 2-8°C Working dilution at room temperature</td>
<td>See expiry date 2 weeks 1 week</td>
</tr>
<tr>
<td>Substrate</td>
<td>Unopened / after opening at 2-8°C</td>
<td>See expiry date</td>
</tr>
<tr>
<td>Stopping solution</td>
<td>Unopened / after opening at 2-8°C</td>
<td>See expiry date</td>
</tr>
</tbody>
</table>

7 TEST PROCEDURE SERION ELISA classic

7.1 Evidence of deterioration

Optimum results can only be achieved if the instructions are strictly followed. Only use SERION ELISA classic reagents when using SERION ELISA classic immunoassays. The components must not be exchanged for reagents of other manufacturers. Standard and control sera of SERION ELISA classic immunoassays are defined exclusively for the test kit to be used and must not be used in other lots. Washing solution, substrate, and stop solution can be used for all SERION ELISA classic immunoassays irrespective of the lot and the test.

Each SERION ELISA classic test contains a ready-to-use sample dilution buffer. In some cases the use of special dilution buffers is necessary to guarantee consistent quality and reliable results. The dilution buffers can be used irrespective of the lots.

There are three different conjugate concentrations for each immunoglobulin class (IgA, IgG, IgM) indicated on the label as + (low), ++ (medium), and +++ (high). Conjugates with the same concentration and of the same immunoglobulin class are interchangeable and can be used for other SERION ELISA classic immunoassays irrespective of the lot and the test. Dilution or alteration of the reagents may result in a loss of sensitivity. Use aseptic techniques when removing aliquots from the reagent tubes to avoid contamination.

Reproducibility of test results is dependent on thorough mixing of the reagents. Agitate the tubes containing control sera before use and also all samples after dilution (e.g. by using a vortex mixer).

Be sure to pipette carefully and comply with the given incubation times and temperatures. Significant time differences between pipetting the first and last well of the microtiter plate when dispensing samples and control sera, conjugate, or substrate can result in different pre-incubation times which may influence the precision and reproducibility of the results. Avoid exposure of reagents to strong light during storage and incubation.
Adequate washing avoids test unspecificities. Therefore, the washing procedure should be carried out carefully. All of the flat bottom wells should be filled with equal volumes of washing buffer. At the end of the procedure ensure that the wells are free of all washing buffer in order to avoid uncontrolled dilution effects. Avoid foaming!

Reagents must be tightly closed after use to avoid evaporation and contamination. Take care not to mix-up the caps of the bottles and/or vials.

The SERION ELISA classic immunoassay is only valid if the lot-specific validation criteria on the quality control certificate are fulfilled.

7.2 Sample Preparation and Storage

Lipaemic, hemolytic or icteric samples (serum or plasma) should only be tested with caution. Obviously contaminated samples should not be tested. Serum or plasma (EDTA, citrate, heparin) collected according to standard laboratory methods are suitable samples. Samples must not be thermally inactivated.

7.2.1 Dilution of Samples

Before running the test, samples ($V_1$) must be diluted in dilution buffer ($V_2$) as follows:

**SERION ELISA classic Respiratory Syncytial Virus IgA**

\[
V_1 + V_2 = 1:400 \quad \text{add} \quad 10\mu l \quad \text{sample} \\
\quad \text{each to} \quad 1000\mu l \quad \text{dilution buffer} \quad (= 1:100) \\
\quad \text{each to} \quad 50\mu l \quad \text{from the first dilution step} \\
\quad \text{each to} \quad 200\mu l \quad \text{dilution buffer} \quad (= 1:4)
\]

**SERION ELISA classic Respiratory Syncytial Virus IgG**

\[
V_1 + V_2 = 1:2000 \quad \text{add} \quad 10\mu l \quad \text{sample} \\
\quad \text{each to} \quad 1000\mu l \quad \text{dilution buffer} \quad (= 1:100) \\
\quad \text{each to} \quad 50\mu l \quad \text{from the first dilution step} \\
\quad \text{each to} \quad 950\mu l \quad \text{dilution buffer} \quad (= 1:20)
\]

**SERION ELISA classic Respiratory Syncytial Virus IgM**

**Interference by rheumatoid factors**

Rheumatoid factors are autoantibodies mainly of the IgM class which preferably bind to IgG immune complexes. The presence of non-specific IgM-antibodies (rheumatoid factors) can lead to false-positive results in the IgM assay. Furthermore, the possibility exists that weak-binding pathogen-specific IgM antibodies are displaced by stronger-binding IgG antibodies leading to a false-negative IgM result. Therefore it is necessary to pretreat samples with rheumatoid factor-absorbent prior to IgM detection (SERION Rf-Absorbent, Order-No. Z200 (20ml/100 tests). Rf-absorption is performed by incubation of the sample in Rf-dilution buffer for 15 minutes at room temperature or overnight at 4°C. The test procedure is described in a separate instruction manual.

Before running the test, rheumatoid factor-absorbent ($V_1$) must be diluted 1:4 in dilution buffer ($V_2$).

\[
V_1 + V_2 = V_3 \quad (1:4) \quad \text{add} \quad 200ul \quad \text{Rf-absorbent} \\
\quad \text{each to} \quad 800ul \quad \text{dilution buffer}
\]
Samples (V₄) must be diluted in this Rf-dilution buffer (V₃):

<table>
<thead>
<tr>
<th>V₄ + V₃ = 1:100</th>
<th>add</th>
<th>10ul</th>
<th>sample</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>each to</td>
<td>1000ul Rf-dilution buffer</td>
<td></td>
</tr>
</tbody>
</table>

After dilution and before pipetting into the microtiter plate the samples must be mixed thoroughly to prepare a homogenous solution.

7.2.2 Sample Storage

Samples should not be stored for more than 7 days at 2-8°C. Extended storage is possible at ≤ -20°C. Avoid repeated freezing and thawing of samples. Diluted samples can be stored at 2-8°C for one week.

7.3 Preparation of kit reagents

Bring all reagents to room temperature before using.

7.3.1 Microtiter Test Strips

The microtiter test strips labeled with abbreviations for pathogen and immunoglobulin class are packed with a desiccant in an aluminum bag. To open the aluminum bag of the microtiter plate please cut off the top of the marked side only in order to guarantee proper resealing. Take unrequired wells out of the frame and put them back into the aluminum bag. Close bag carefully to ensure airtight conditions. Do no use the strips if the aluminum bag is damaged or if the bag with remaining strips and desiccant was not properly resealed.

7.3.2 Control Sera / Standard Sera (ready-to-use)

Control and standard sera are ready-to-use and must not be diluted any further. For each test run - independent of the number of microtest strips to be used - control and standard sera must be included. Standard and cut-off sera should be set up in duplicate. Do not treat control sera with Rf-absorbent.

7.3.3 Anti-human-IgA, IgG, or IgM AP-Conjugate (ready-to-use)

The required conjugate concentration (+, ++, ++++) is indicated on the quality control certificate. Please refer also to the specification on the label.

7.3.4 Washing Solution (Concentrate)

Dilute washing buffer concentrate (V₁) 1:30 with distilled water to a final volume of V₂.

Example:

<table>
<thead>
<tr>
<th>Buffer concentrate (V₁)</th>
<th>Final volume (V₂)</th>
</tr>
</thead>
<tbody>
<tr>
<td>33.3 ml</td>
<td>1000 ml</td>
</tr>
<tr>
<td>1 ml</td>
<td>30 ml</td>
</tr>
</tbody>
</table>

7.3.5 Dilution Buffer for Samples (ready-to-use)

7.3.6 Substrate (ready-to-use)

Substrate in unopened bottle may have a slight yellow color which does not reduce the quality of the product!

7.3.7 Stopping Solution (ready-to-use)
7.4 Overview - Test Procedure

SERION ELISA \textit{classic}
Respiratory Syncytial Virus (RSV) IgA/IgG quantitative
Respiratory Syncytial Virus (RSV) IgM qualitative

sample dilution$^1$

IgA: 1:500
IgG: 1:2000
IgM: 1:100

Pipette diluted samples and ready-to-use control / standard sera into the microtest wells (100µl)

$\text{INCUBATION \ 60 \min. / \ 37^\circ C}$

moist chamber

$\downarrow$

WASH (4 x 300ul DIL WASH)$^2$

Pipette conjugate solution (100µl)

$\text{INCUBATION \ 30 \min. / \ 37^\circ C}$

moist chamber

$\downarrow$

WASH (4 x 300ul DIL WASH)$^2$

Pipette substrate solution pNPP (100µl)

$\text{INCUBATION \ 30 \min. / \ 37^\circ C}$

moist chamber

$\downarrow$

Pipette stopping solution STOP (100µl)

READ EXTINCTION AT 405 nm

$^1$Special dilution buffers for the following SERION ELISA \textit{classic} tests: Borrelia burgdorferi IgG, IgM, EBV EA IgG and Hantavirus Puumala IgG, IgM

$^2$For manual use:
tap plate at the end of the wash procedure on paper towel
7.5 Manual Test Procedure

1. Place the required number of cavities in the frame and prepare a protocol sheet.
2. Add each **100 µl of diluted sample or ready-to-use controls** into the appropriate wells of microtest strips. Spare one well for substrate blank, e.g.:

<table>
<thead>
<tr>
<th>Well</th>
<th>Quantitative ELISA</th>
<th>Qualitative ELISA</th>
</tr>
</thead>
<tbody>
<tr>
<td>A1</td>
<td>Substrate blank</td>
<td>Substrate blank</td>
</tr>
<tr>
<td>B1</td>
<td>Negative control</td>
<td>Negative control</td>
</tr>
<tr>
<td>C1</td>
<td>Standard serum</td>
<td>Cut-off serum</td>
</tr>
<tr>
<td>D1</td>
<td>Standard serum</td>
<td>Cut-off serum</td>
</tr>
<tr>
<td>E1</td>
<td>Sample 1 . . .</td>
<td>Positive control</td>
</tr>
<tr>
<td>F1</td>
<td>Sample 2 . . .</td>
<td>Sample 1 . . .</td>
</tr>
</tbody>
</table>

3. **Sample incubation** for 60 minutes (+/- 5 min) at 37°C (+/- 1°C) in moist chamber.
4. After incubation **wash** all wells with washing solution (by automated washer or manually):
   - aspirate or shake out the incubation solution
   - fill each well with 300µl washing solution
   - aspirate or shake out the washing buffer
   - repeat the washing procedure 3 times (altogether 4 times!)
   - dry by tapping the microtest plate on a paper towel
5. **Addition of conjugate**
   - Add 100µl of the ready-to-use IgA/IgG/IgM conjugate to the appropriate wells (except substrate blank)
6. **Conjugate incubation** for 30 minutes (+/- 1 min) at 37°C (+/- 1°C) in moist chamber.
7. After incubation **wash** all wells with washing solution (see above)
8. **Addition of substrate**
   - Add 100µl of ready-to-use substrate solution to each well (including well for substrate blank!)
9. **Substrate incubation** for 30 minutes (+/- 1 min) at 37°C (+/- 1°C) in moist chamber.
10. **Stopping of the reaction**
    - Add 100µl stopping solution to each well, shake microtest plate gently to mix.
11. **Read optical density**
    - Read optical density (OD) within 60 minutes at 405nm against substrate blank, reference wave length between 620nm and 690nm (e.g. 650nm).

7.6 Automated Test Procedure

SERION ELISA are suited for processing on automats and evaluated for use with Immunomat™ and Gemini as well as with DYNEX DSX® and DS2®. The automated processing is performed analogous to manual use. Please note that under special working conditions (e.g. ambient temperature) internal laboratory adaptations of the substrate incubation times may be necessary.

7.7 Positive Control / Accuracy Control

For the periodic verification of the test method, in order to fulfill the requirements of laboratory internal quality management systems, we recommend using SERION ELISA controls to determine precision and accuracy of SERION ELISA classic test runs. The use of SERION ELISA controls is described in specific instruction manuals.
8. TEST EVALUATION

8.1 SERION ELISA classic Respiratory Syncytial Virus (RSV) IgA/IgG

The mathematical curve fitting for antibody quantification with SERION ELISA classic immunoassays is based on the 4-parameter logistic (4PL) function.

\[ \text{Activity} \left( \frac{U}{ml} \right) = e^{\frac{1}{B} \ln\left( \frac{D-A}{OD(\text{Patient}+F-A)} \right)} \]

The 4 parameters A, B, C, and D are representative for the exact shape of the curve:

- Parameter A: Lower asymptote (OD)
- Parameter B: Slope of the curve
- Parameter C: Inflection point
- Parameter D: Upper asymptote (OD)

Institut Virion\Serion GmbH establishes a lot-specific 4PL standard curve for each SERION ELISA classic immunoassay in multiple test runs under optimal test conditions. The four parameters are indicated on the quality control certificate of each individual SERION ELISA classic test.

For the adaptation of the test level to the given 4PL standard curve the correction factor \( F \) is calculated by dividing the standard reference OD value indicated on the quality control certificate with the measured, and consequently test run-specific, standard OD value.

\[ F = \frac{\text{STD reference OD value}}{\text{measured \ STD \ OD \ value}} \]

By multiplying the OD values obtained from samples with the correction factor \( F \), the level of each individual test run is adjusted to the given 4PL standard curve. Thereby, interassay deviations are compensated for and antibody activities can be directly evaluated from the 4PL standard curve.

After subtraction of the substrate blank from all measured OD values and calculation of the mean OD value of the standard serum (STD), tested in duplicate, a range of possibilities are available for the evaluation of antibody activities from the optical measurement signals (OD) of samples. They are described in separate manuals.

8.2 Borderline Ranges

The borderline ranges of the SERION ELISA classic Respiratory Syncytial Virus (RSV) IgA/IgG tests are specified on the quality control certificates and indicate the range of borderline test results. Values below this range indicate a negative test result; values above the borderline range indicate a positive test result.

8.3 Limits of Quantification

The limits of quantification are specified on the quality control certificate of the SERION ELISA classic Respiratory Syncytial Virus (RSV) IgA/IgG. The linearity of dilution within this range has been demonstrated in comprehensive evaluation studies. If a sample shows a test result above the upper limit of quantification, the sample may be tested at a higher dilution. The resulting antibody activity must then be multiplied by the additional dilution factor.
8.4 SERION ELISA classic Respiratory Syncytial Virus (RSV) IgM

For the evaluation of test runs a lot-specific quality control certificate with declarations concerning cut-off serum and positive control is included in every SERION ELISA classic.

Before evaluation the blank value (blank) has to be subtracted from each sample value. For determination of the cut-off range in OD the mean of the readings for the cut-off serum has to be calculated. The cut-off range in OD corresponds to the mean value of the cut-off serum +/- 10%.

<table>
<thead>
<tr>
<th>OD sample</th>
<th>more than 10% over</th>
<th>OD cut-off</th>
<th>positive</th>
</tr>
</thead>
<tbody>
<tr>
<td>OD sample</td>
<td>+/- 10% of</td>
<td>OD cut-off</td>
<td>borderline</td>
</tr>
<tr>
<td>OD sample</td>
<td>more than 10% under</td>
<td>OC cut-off</td>
<td>negative</td>
</tr>
</tbody>
</table>

8.5 Automated Evaluation / Software

For the automated evaluation of optical measurement signals, the software SERION easyANALYZE, the software SERION evaluate as well as the Microsoft® Excel®-based software tool SERION activity are available on request.

8.6 Criteria of Validity

- The substrate blank must be <0.25 OD.
- The negative control must be negative.
- By use of quantitative SERION ELISA classic tests the mean OD value (after subtraction of the substrate blank!) of the standard serum must be within the validity range which is given on the lot-specific quality control certificate.
- By use of qualitative SERION ELISA classic tests the OD value of the positive control and the mean OD value of the cut-off serum must be within the validity ranges which are given on the lot-specific quality control certificate of the kit (after subtraction of the substrate blank!)
- The variation of OD values of the standard serum or cut-off serum must not be higher than 20%.

If these criteria are not met, the test is not valid and must be repeated.

9. SAFETY MEASURES

9.1 Statements of Warning

The SERION ELISA classic is designed for use by qualified personnel who are familiar with good laboratory practice. All kit reagents and samples should be handled carefully using established good laboratory practice.

- This kit contains human blood components. Although all control and cut-off sera have been tested and found negative for anti-HIV Ab, HBs-Ag (Hepatitis B Virus surface-Antigen) and anti-HCV-Ab, they should be considered potentially infectious.
- Do not pipette by mouth.
- Do not smoke, eat, or drink in areas in which samples or kit reagents are handled.
- Wear disposable glove, laboratory coat and safety glasses while handling kit reagents or samples.
- Wash hands thoroughly afterwards.
- Samples and other potentially infectious material should be decontaminated after the test run.
- Reagents should be stored safely and be inaccessible to unauthorized access, e.g. children.

9.2 Disposal

Please observe the relevant statutory requirements!